Research Article [Araştırma Makalesi]

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# The effects of dexmedetomidine on oxidant - antioxidant systems in the experimental model of carbondioxide pneumoperitoneum

[Deneysel karbondioksit pnömoperitonyum modelinde deksmedetomidin'in oksidan-antioksidan sistem üzerine etkisinin araştırılması]

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#### ABSTRACT

**Objective:** The aim of the study was to investigate the changes of oxidative and anti-oxidative systems in the splanchnic area during carbon dioxide pneumoperitoneum and to determine whether the administration of dexmedetomidine has effects on these systems.

**Methods:** Forty rats were randomized into four groups: Group I; Control, Group II; No pneumoperitoneum, Dexmedetomidine administration, Group III; Pneumoperitoneum, no Dexmedetomidine administration and Group IV; Pneumoperitoneum and Dexmedetomidine administration 30 minutes before insufflation. The rats were rested 30 minutes after desufflation and blood samples were obtained for; ischaemia modified albumin (IMA), myeloperoxidase (MPO), advanced oxidation protein products (AOPP), catalase (CAT), paraoxonase (PON1) and platelet-activating factor acetylhydrolase (PAF-AH) analyses.

**Results:** When compared with the control group; the serum IMA levels significantly decreased in group II, and also increased in group III as compared to control (p<0.05). IMA levels were also significantly decreased in both groups II and IV as compared to group III (p<0.001). Serum MPO activity increased in group III as compared to control (p<0.05). Serum AOPP levels were significantly increased in group III as compared to group II (p<0.01) and decreased in group IV as compared to group II (p<0.01). Serum CAT activity was higher in group II than controls (p<0.05). Serum PON and plasma PAF-AH activities significantly decreased in group III as compared to group II (p<0.05) and plasma PAF-AH activity were decreased in group III as compared to controls (p<0.05).

**Conclusion:** In conclusion, administration of dexmedetomidine; prior to ischemia reperfusion injury caused by pneumoperitoneum; reduces the oxidative injury and increases the antioxidant activity in the acute period.

Key Words: Dexmedetomidine, ischemia-reperfusion injury, oxidant-antioxidant system, pneumoperitoneum.

**Conflict of Interest:** The authors declare no conflict of interest.

## ÖZET

Amaç: Bu çalışmanın amacı, karbondioksit pnömoperitonyum sırasında splanknik sahada oluşan oksidan ve antioksidan sistemlerdeki değişiklikleri ve deksmedetomidin uygulamasının bu değişiklikler üzerine etki gösterip göstermediğini tespit etmektir.

**Metod:** Kırk adet rat randomize olarak dört gruba ayrıldı. Grup I; kontrol, Group II; pnömoperitonyum uygulanmayan sadece deksmedetomidin verilen grup, Group III; pnömoperitonyum uygulanan ancak deksmedetomidin verilmeyen grup ve Grup IV; pnömoperitonyum uygulanan ve insuflasyondan 30 dakika önce deksmedetomidin verilen grup. Desuflasyon sonrası 30 dakika dinlendirilen ratlardan; iskemi modifiye albümin (İMA), miyeloperoksidaz (MPO), ileri oksidasyon protein ürünleri (AOPP), katalaz (CAT), paroksonaz (PON1) ve trombosit aktive edici faktör asetil hidrolaz (PAF-AH) analizi için kardiyak ponksiyonla kan örnekleri alındı.

**Bulgular:** Kontrol grubu ile karşılaştırıldığında, serum İMA seviyeleri Grup II'de azalmış Grup III'de ise artmış olarak bulundu (p<0.05). Grup III ile karşılaştırıldığında ise serum İMA seviyeleri Grup II ve IV'de azalmış olarak bulundu (p<0.001). Kontrol grubu ile karşılaştırıldığında serum MPO aktivitesi Grup III'de artmış olarak tespit edildi (p<0.05). Serum AOPP seviyeleri Grup II ile karşılaştırıldığında Grup III'de artmış olarak tespit edildi (p<0.05). Serum AOPP seviyeleri Grup II ile karşılaştırıldığında Grup III'de artmış olarak (p<0.01), Grup IV'de ise Grup III'e göre azalmış olarak tespit edildi (p<0.05). Serum AOPP seviyeleri Grup II'e göre azalmış olarak tespit edildi (p<0.05). Serum PON ve plazma PAF-AH aktivitesi Grup II'ye göre Grup III'de anlamlı olarak düşük (p<0.05) ayrıca plazma PAF-AH aktivitesi kontrol grubuna göre Grup III'de de anlamlı olarak düşük bulundu (p<0.05).

**Sonuç:** Pnömoperitonyum neden olduğu iskemi reperfüzyon hasarı öncesinde deksmedetomidin verilmesinin, akut dönemde oksidatif hasarı azaltıp, antioksidan aktiviteyi artırarak olumlu etkiye neden olabileceği gösterilmektedir.

Anahtar Kelimeler: Deksmedetomidin, iskemi-reperfüzyon hasarı, oksidan-antioksidan sistem, pnömoperitonyum.

Çıkar Çatışması: Yazarların çıkar çatışması yoktur.

# Introduction

The insufflation of the abdomen with carbondioxide to provide the required space for laparoscopic surgery is named as carbon dioxide pneumoperitoneum (CO, Pp). The periods of insufflation and desufflation constitutes a typical ischemia reperfusion injury (I/R injury) in the splanchnic area during this procedure. Today, laparoscopic surgery has become the gold standard for many procedures over the past decade but it is also accompanied by I/R injury as a result of CO, Pp. Hypoperfusion of the splanchnic area and the I/R injury, precipitate to release free radicals. It is known that according to injury of these radicals to the cell membrane, tissue integrity is damaged and the antioxidant capacity of the organism is reduced [1-3]. The laparoscopic procedures with possible I/R injury and major abdominal surgery especially accompanying sepsis, serious trauma are very important acute inflammation cascades and they require meticulous management of anesthesia [4].

It is known that some type of anesthesics have protective potency from I/R injury [5]. Dexmedetomidine (DEX) is an adjuvant anesthesic drug that has sedative, hypnotic and analgesic potency. DEX can be used as an adjuvant anesthesic drug and some experimental and clinical studies demonstrated that DEX has protective effects against injury in the oxidative stress since it attenuates the inflammatory response [6-11]. There are various biochemical markers that indicative of oxidative stress and antioxidant defense system and which were also used to investigate the effects of DEX against I/R injury [12-16].

Ischaemia modified albumin (IMA) is the metabolic variant of albumin. The N-terminal amine group of human albumin is the metal binding part of the protein that binds ions like cobalt, nikel, and copper in plasma. The superoxide anions and the other reactive oxygen molecules which appears during acidosis, ischemic and oxidative reactions make changes on the structure of N-terminal and finally a re-constituted metabolic variant of albumin takes place in plasma [9,17]. Myeloperoxidase (MPO) is a lysozome enzyme that appears as a response to oxidative stress [18]. Advanced oxidation protein products (AOPP) occurs as a result of aggregation and fragmentation of proteins due to reactive oxygen radicals. AOPP is a new marker that shows the effect of oxidative stress on proteins in inflammation and widely used in many studies as an indicator of I/R injury [18,19]. Toxic oxygen radicals and its derivates which appear during I/R injury are cleared by free radical cleaner such as catalase (CAT) that is an antioxidant, enzymatic defender already exists in the metabolism naturally [20,21]. Paraoxonase (PON1) is an ester hydrolase glycoprotein that has arylesterase, paraoxonase and lactonase activity [22,23]. Platelet-activating factor acetylhydrolase (PAF-AH) is a pro-inflammatory molecule which has anti-oxidant and anti-inflammatory properties that protects metabolism by hydrolysing platelet activating factor (PAF) and oxidated phospholipids.

According to other routine markers PAF-AH is relatively new biochemical marker [24].

The main hypothesis of the present experimental study is based on whether DEX, an adjuvant anesthetic drug, administration has a positive impact on I/R injury in CO<sub>2</sub> Pp model in rats using the biochemical markers such as: IMA, MPO, AOPP and antioxidant enzymes; CAT, PON1 and PAF-AH.

## **Materials and Methods**

## Animals

This experimental study was approved by Ondokuzmayıs University Animal Care and Ethics Committee (2013/06). Female Sprague–Dawley rats (n: 40, weight: 200–250g) were randomized into four groups. There was no mortality in the study animals and all rats were stable throughout the perioperative period. Rats were anesthetized with ketamine hydrochloride (i.m. 60 mg/kg) and xylazine (i.m. 10 mg/kg) and anesthesia was maintained with xylazine (i.m.10 mg/kg per hour). Group I; Control group, Group II; No CO, Pp, DEX administrated in dose 100 µg/kg (Precedex 200µg/2ml, Hospira Inc, USA) i.p., Group III; CO<sub>2</sub> Pp with intra abdominal pressure of 12 mmHg for 60 minutes, no DEX administration and Group IV; CO, Pp with intra abdominal pressure of 12 mmHg for 60 minutes and DEX administrated in dose 100 µg/kg i.p. 30 minutes before insufflation of CO, Pp. In Group I and III, an equal volume of serum physiologic with Dexmedetomidine, was administered i.p. The rats were placed in supine position with their extremities to the operating table. CO<sub>2</sub> Pp was achieved by peritoneal cavity puncture with an 18 G cannula, placed caudally to the sternum and CO<sub>2</sub> was insufflated by this canulla with Karl Storz insufflator in animal research laboratory. Pp was performed by insufflation of CO<sub>2</sub> to maximum intra abdominal pressure of 12 mmHg and intraabdominal pressure was checked over 60 minutes with this insufflator. The rats were rested 30 minutes after desufflation and each of 6 ml blood samples were obtained for; IMA, MPO, AOPP, CAT, PON1 and PAF-AH analyses by cardiac puncture. After this procedure the animals were sacrified by exsanguination. The blood samples were centrifuged at 2000 X g for 10 min at 4°C and serum samples were frozen at -70 °C for two months.

## **Biochemical analyses**

#### Serum IMA Assay

Levels of IMA were measured using the commercial Enzyme-Linked Immunosorbent Assay (ELISA) kit (Cusabio Biotech Wuhan, China) with detectable IMA ranging from 7.8 to 500 ng/mL.

#### Serum MPO Assay

Serum MPO activity was determined by the method of Bradley et al. and was based on kinetic measurement of the formation rate of the yellowish-orange product of the oxidation of o-dianisidine with MPO in the presence of  $H_2O_2$  at 460 nm. One unit of MPO was defined as that degrading 1 µmol of  $H_2O_2$  per minute at 25°C. A molar extinction coefficient of 1.13 x 10<sup>4</sup> M<sup>-1</sup>cm<sup>-1</sup> of oxidized o-dianisidine was used for the calibration. MPO activity was expressed in units per litre of serum (U/L) [25].

#### Serum AOPP Assay

Determination of AOPP was based on a spectrophometric assay according to Witko-Sarsat et al. One millilitre of serum diluted 1:5 with phosphate-buffered saline (PBS, pH7,4), 1 ml of chloramine-T (0-100  $\mu$ mol/L) for calibration and 1 ml of PBS as blank were placed on corresponding tubes; 50  $\mu$ L of 1.16 M potassium iodide (KI) was then added to each tube, and 2 min later 200  $\mu$ L glacial aceticacid was added. The absorbance of there action mixture was read immediately at 340 nm. AOPP level was expressed in  $\mu$ mol of chloramine-T equivalents per litre of serum ( $\mu$ mol/L) [26].

## Serum CAT Assay

The serum CAT activity was determined by Goth's colorimetric method in which serum samples were incubated with  $H_2O_2$  substrate and the enzyme reaction were stopped by the addition of ammonium molybdate. The intensity of the yellow complex formed by molybdate and  $H_2O_2$  was measured at 405 nm. Serum CAT activity was expressed as U/L [27].

## Serum PON1 Assay

Paraoxonase activity was measured according to Gan et al. using phenylacetate as substrate at a final concentration of 1 mM. Formation of phenol at 25°C was monitored at 270 nm, in the presence of 50 mM Tris-HCl (pH 8) containing 1 mM CaCl<sub>2</sub> and 40  $\mu$ M eserine. Enzymatic activity was calculated from the molar extinction coefficient of phenol ( $C_{270} = 1310 \text{ M}^{-1}\text{cm}^{-1}$ ) and corrected for the non-enzymatic hydrolysis. One unit of paraoxonase activity is defined as 1  $\mu$ mol of substrate hydrolyzed permin, under the defined assay conditions [28].

## Serum PAF-AH Assay

PAF-AH activity was determined spectrophotometrically using a modification of the method described by Stafforini et al [29]. 2-thio-PAF (Cayman Chemical) was used as substrate at a final concentration of 1 mM in 100 mM Tris-HCl buffer (pH 7,4). Hydrolysis of thioester substrate was measured at 37°C, using Ellman's procedure for monitoring at 412 nm the accumulation of free sulfhydryl groups reacting with DTNB. Enzymatic activity was calculated from the molar extinction coefficient of DTNB ( $\varepsilon_{412}$ = 13600 M<sup>-1</sup>cm<sup>-1</sup>). One unit of PAF-AH activity is defined as 1 µmol of substrate hydrolyzed per min, under the defined assay conditions.

## Statistical analyses

The data are expressed as mean  $(X) \pm$  Standart deviation

|               | Results of Kolmogorov-Smirnov test |         | Results of ANOVA |         |
|---------------|------------------------------------|---------|------------------|---------|
|               | KS statistics                      | p value | F Statistics     | p value |
| IMA (ng/mL)   | 0.120                              | 0.157   | 14.092           | 0.001   |
| MPO (U/L)     | 0.140                              | 0.075   | 4.437            | 0.010   |
| AOPP (µmol/L) | 0.143                              | 0.052   | 8.693            | 0.001   |
| CAT (U/L)     | 0.139                              | 0.072   | 3.236            | 0.035   |
| PAF-AH (U/mL) | 0.075                              | 0.150   | 5.799            | 0.003   |
| PON1(U/mL)    | 0.074                              | 0.152   | 2.833            | 0.050   |

Table 1. K-S and ANOVA results for the measured parameters

|  | Table 2. | Descriptive statistics | and comparison results | for the measured parameters |
|--|----------|------------------------|------------------------|-----------------------------|
|--|----------|------------------------|------------------------|-----------------------------|

|               | Control<br>x±SD | Group II<br>x±SD             | Group III<br>x±SD              | Group IV<br>x±SD             |
|---------------|-----------------|------------------------------|--------------------------------|------------------------------|
| IMA (ng/mL)   | 3.24±0.27       | 2.90±0.26 <sup>a*. c**</sup> | 3.54±0.21ª*                    | 3.05±0.13 <sup>c**</sup>     |
| MPO (U/L)     | 127.32±57.08    | 312.90±175.19                | 380.78±212.91ª*                | 266.45±118.08                |
| AOPP (µmol/L) | 298.84±63.54    | 205.75±71.49                 | 512.9±233.39 <sup>b**</sup>    | 272.26±104.01 <sup>c**</sup> |
| CAT (U/L)     | 111.18±41.15    | 229.20±96.42 <sup>a*</sup>   | 162.72±60.27                   | 226.87±147.98                |
| PAF-AH (U/mL) | 0.1332±0.039    | 0.1358±0.0045                | 0.1269±0.0055 <sup>a*.b*</sup> | 0.1315±0.0050                |
| PON1(U/mL)    | 125.90±5.82     | 129.22± 3.99                 | 123.90±2.92 <sup>b*</sup>      | 126.60±2.45                  |

\*: According to Tukey test. different lower cases represent statistically significant differences among the groups. a; compared to control group. b; compared to group II. c; compared to group III. \*;p<0.05. \*\*p<0.01

Table 3. The relationship between all parameters in group II

| Parameters | MPO   | AOPP  | CAT   | PON1  | PAFAH |
|------------|-------|-------|-------|-------|-------|
| AOPP       | 0.78* | 1     |       |       |       |
| CAT        | 0.26  | -0.03 | 1     |       |       |
| PON1       | -0.42 | -0.21 | -0.24 | 1     |       |
| PAFAH      | -0.43 | 0.12  | -0.23 | 0.23  | 1     |
| IMA        | 0.34  | 0.65  | -0.40 | -0.20 | 0.71* |
|            |       |       |       |       |       |

\*: p<0.05

Table 4. The relationship between all parameters in group III

| MPO   | AOPP                          | CAT   | PON1  | PAFAH  |
|-------|-------------------------------|---|---|--|
| 0.21  | 1                             |   |   |  |
| 0.66* | 0.54                          | 1   |   |  |
| 0.26  | 0.06                          | -0.87                                       | 1   |  |
| 0.28  | 0.09                          | -0.03                                       | 0.18  | 1  |
| -0.29 | -0.45                         | -0.35                                       | -0.53   | 0.70   |
|       | 0.21<br>0.66*<br>0.26<br>0.28 | 0.21 1   0.66* 0.54   0.26 0.06   0.28 0.09 | 0.21 1   0.66* 0.54 1   0.26 0.06 -0.87   0.28 0.09 -0.03 | 0.21   1     0.66*   0.54   1     0.26   0.06   -0.87   1     0.28   0.09   -0.03   0.18 |

\*: p<0.05

Table 5. The relationship between all parameters in group IV

| Parameters | MPO    | AOPP  | CAT  | PON1  | PAFAH |
|------------|--------|-------|------|-------|-------|
| AOPP       | 0.10   | 1     |      |       |       |
| CAT        | 0.90** | 0.24  | 1    |       |       |
| PON1       | 0.54   | 0.61* | 0.21 | 1     |       |
| PAFAH      | 0.40   | 0.19  | 0.28 | -0.22 | 1     |
| IMA        | 0.33   | 0.25  | 0.09 | 0.23  | 0.33  |
|            |        |       |      |       |       |

\*: p<0.05, \*\*: p<0.01

(SD). Kolmogrov-Smirnov (K-S) Goodness of Fit Test was used to control whether the distribution of parameters was normal or not. Then groups of data were compared with an analysis of variance (One-way ANOVA) followed by Tukey's multiple comparison tests. In addition, Pearson correlation analysis was carried out for determination of linear relationships among the variables. Statistical significance was considered as 5% for all statistical computations.

## Results

Kolmogrov-Smirnov (K-S) Goodness of Fit Test and ANOVA results for for all variables are presented in Table 1. The comparisons of measured parameters are also presented in Table 2. As shown in Table 2; the serum IMA level significantly decreased in group II, and also increased in group III as compared to control (p<0.05). However, IMA levels were significantly decreased in both groups II and IV as compared to group III (p<0.01). Significantly increased serum MPO activity was obtained in group III as compared to controls (p<0.05). Serum AOPP levels were increased significantly in group III as compared to group II (p<0.01) and significantly decreased in group IV compared to group III (p<0.01). The serum CAT activity was higher in group II than control (p<0.05). Serum PON1 activity significantly decreased in group III as compared to group II (p<0.05). Serum PAF-AH activity were decreased in group III as compared to control and group II (p < 0.05). There were no significantly differences among the other parameters in all groups (p>0.05). The correlations between all parameters in Group II, III and IV are presented in Table 3, 4 and 5. There were significantly correlation between MPO and AOPP (p<0.05, r=0.78), PAF-AH and IMA (p<0.05, r=0.71) in group II, MPO and CAT (p<0.05, r=0.66) in group III, and also MPO and CAT (p<0.01, r=0.90), PON1 and AOPP (p<0.05, r=0.61) in group IV.

## Discussion

I/R injury precipitate to release free radicals. It is known that according to injury of these radicals to the cell membrane, tissue integrity is damaged and the antioxidant ca-

pacity of the organism is reduced [1,2]. It is demonstrated in human and experimental studies that to establish Pp for laparoscopic procedures causes I/R injury [3,20]. Although laparoscopy is a well-tolerated procedure by patients, oxidative stress caused by  $CO_2$  Pp may result in co-morbidity [20].

It is proven with previous studies that, type of anesthesia and the agent used in the procedure associates with oxidative stress, so they play an active role in I/R injury [5,30]. DEX is a potent and specific  $\alpha_2$  adrenoceptor agonist that reduces proapopitotoic proteins via  $\alpha_2$  adrenoceptor stimulation. Sedative and analgesic features of DEX provide this drug as an adjuvant therapy for perioprative period. Moreover it is known that, DEX reduces mortality by reducing the level of TNF $\alpha$  and increases the functions of macrophages especially phagocytose and finally reduces proinflammatory cytokines and inflammatory response [6,10].

IMA is an early and sensitive marker of myocardial, pulmonary, mesenteric and cerebral ischemic processes that indicates the oxidative stress [9,17]. IMA was demonstrated as a useful marker that shows ischemic changes in splanchnic area during Pp in previous studies [31]. It is observed that IMA levels were increased in only Pp group and also decreased in Pp with DEX administration group in our study. Previous studies conclude that DEX is a useful drug in I/R injury by preventing lipid peroxidation [32,33]. Recently Geze et al. showed that DEX administration before Pp caused to lower IMA levels in their experimental study and they concluded that DEX prophylaxis reduced the post-pneumoperitoneum I/R injury [9]. It was observed in a histopathological study on ovarian tissue of rats with Pp that DEX administration resulted in decrease for vascular congestion, hemorrhage and follicular degeneration. Moreover, it was also reported that DEX reduces oxidative stress in the same study [34]. DEX administration resulted in significantly decreasing level of IMA and increasing CAT activity when compared to control group. It was also observed statistically significant correlation between IMA levels and PAF-AH activity in the group II. We believe that antioxidant effect of DEX derives from increasing CAT activity. Acute inflammatory response secondary to neutrophil activation in I/R injury releases reactive oxygen molecules and cytotoxic proteins such as MPO from neutrophils to the extracellular fluid [35]. An experimental study reported by Zhang et al. showed that an intestinal I/R injury by clemping superior mesenteric artery shows that MPO levels are increasing in I/R injury [30]. It is reported that MPO increases during the ischemic period due to Pp and the reperfusion period after decompression [35]. When compared with the control group, MPO activity was higher in the Pp only group and it was thought that it increases as a result of I/R injury in our study. The levels of MPO in DEX administration before Pp group were lower but it was not statistically

significant. MPO is a commonly used enzymatic marker in order to show the level of inflammation. Uysal et al. reported that flap necrosis associates with high MPO levels and they stated that inflammatory response and MPO levels significantly decrease in DEX administration group [7]. Kontoulis et al. showed that Pp induced I/R injury causes higher MPO levels and also stated that long duration of Pp and reperfusion causes more increseasing in MPO levels [36]. MPO activities in our study were measured in the blood samples that were taken 60 minutes after Pp. We consider the difference between the results of our study and the others originate from the timing of the bood sampling. For that reason we think that further studies are needed to investigate the protective effects of DEX during pneumoperitoneum with different dose regimens in late period of the injury.

Measurement of serum AOPP level may be a useful marker of protein oxidative damage [16]. It is demonstrated that I/R injury secondary to Pp in laparoscopic surgeries increases AOPP levels in a previous study [37]. In the only DEX administration group, there was statistically significant correlation between MPO and AOPP which are the markers of I/R injury. Likewise, AOPP levels in Pp only group were found to be increased in our study. The decrease in AOPP level in DEX administration before Pp group was thought to be a result of its anti-inflammatory effect.

Toxic oxygen radicals and its derivates which appear during I/R injury are cleared by free radical cleaner such as CAT. It degrades hydrogen peroxide into water and oxygen [20]. Bulbuloglu et al. reported in an experimental study that CAT activity decrease in I/R injury secondary to Pp and increases after administration of antioxidant [38]. An experimental model of I/R injury of muscle tissue which assess antioxidant activities with DEX administration, CAT activity in I/R injury group was found lower, on the other hand CAT activity in DEX administration group before reperfusion was higher than control group. Dong et al. concludes that DEX has protective effects in I/R injury [39]. In our present study, significantly higher CAT activity was observed in only DEX administration group as compared with control group. Although it was not statistically significant, a relative increase of CAT activity in DEX administration before Pp group would be accepted as a positive effect of DEX administration on antioxidant system. In this study, there was statistically significant correlation between MPO and CAT activity in Group III and IV. MPO and CAT are the enzymes that use hydrogen peroxide radicals as a substrate. Although CAT activity in Group III and IV was not significantly higher than control group, MPO activity in these groups was significantly higher than control group. We believe that obtained significant correlation between MPO and CAT activity is related to increasing level of hydrogen peroxide secondary to Pp.

PON1, that decrease in inflammation and oxidative stress is used as an indicator of antioxidative capacity [23]. An experimental study results showed that DEX administration before I/R injury caused to decrease total antioxidant capacity and PON1 activity in rats with hepatic I/R injury [8]. In our present study PON1 levels in Pp only group were significantly lower than DEX administration without Pp group. This result proves that Pp results in I/R injury. PON1 levels in DEX administration before Pp group were not statistically significant but it shows that DEX administration makes PON1 levels closer to control group. DEX administration caused to significant decrease on levels of AOPP and PON1 activity in Group IV. Although DEX administration didn't cause to increase on the antioxidant enzyme activity in group IV, there was also statistically significant correlation between AOPP and PON1. The results of the present study demonstrate that DEX protects the metabolism by attenuating the oxidative stress. In the presence of ischemia and inflammation, PAF-AH is inactivated by oxygen radicals in the circulation irreversibly [14]. The level of PAF-AH is low in patients with oxidative stress based diseases. The examples of these disesases are; asthma, systemic lupus erythematosus, acute myocard infarction, multiple organ failure and sepsis [40]. In our study, the levels of PAF-AH are found to be significantly low in the Pp only group and this result was an expected result because of its antioxidant nature of this marker which tends to decrease in ischemia. In the review of the literature, there was no study that investigates the effects of DEX on PAF-AH levels in Pp model. Our present study is the first experimental study in this manner. PAF-AH levels were low as it was expected I/R injury during Pp. Even it was not found to be statistically significant, PAF-AH levels in DEX administration before Pp group were close to control group and this result may suggest that DEX administration can tip the balance in favor of antioxidative side.

# Conclusion

Our experimental I/R injury model demonstrated not only the increasing levels of IMA, MPO and AOPP which are oxidative markers but also the decreasing levels of PON1 and PAF-AH activity that have antioxidant potential during pneumoperitoneum. The administration of DEX may minimize the harmful effects of oxidative injury caused by Pp.

## Ethical Issues

This experimental study was approved by Ondokuz Mayıs University Animal Care and Ethics Committee (2013/06), Samsun, Turkey.

## Supporting organizations

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# **Conflict of Interest**

There are no conflicts of interest among the authors.

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